MENOUFIA JOURNAL OF AGRICULTURAL BIOTECHNOLOGY

https://mjab.journals.ekb.eg

CORRELATION BETWEEN QUALITATIVE AND QUANTITATIVE CELLULASE ENZYMES ACTIVITIES IN SOME *TRICHODERMA SPP*.

El-Sobky M. A.*; Abdel-Lateif, K.S.; Fahmi, A. I.; El-Zanaty, A. M. and Eissa Ragaa. A.

Genetics Department, Faculty of Agriculture, Menoufia University, Shebin El-Kom, Egypt

Received: Feb. 15, 2024 Accepted: Feb. 29, 2024

ABSTRACT: Twenty-six isolates used in qualitative screening of Cellulase done in two media CMC-Congo red dye plate and 1% PASC (phosphoric acid swollen cellulose) plate. Quantitative Enzymes activities were tested in two fermentation methods SSF (solid state fermentation) and SmF (Submerged fermentation). SmF was detected at 2,3,4,6 and 7 days in Mandel's Media with CMC 0.5%. Moreover, SSF was carried out by using straws (Rice and wheat) for all isolates. The cellulase activities measured as (FPase, CMCase and β -glucosidase). In this study positive correlation coefficient in enzymes activity between SSF and qualitative screening in (CMC & PASC) plate. Higher correlation have been detected between PACS and SSF rice straw was about (0.595**) for β -Glucosidase. After all these tests we can compare enzyme production by SmF and SSF. Data showed SmF production for high yield and high quality. The cellulase genes, *Chb* gene encoding cellobiohydrolase 2 (CBH2) and *Bgl* gene that encoding endo- β gluconase (BGL1) were present in all *Trichoderma* isolates.

Key words: *Trichoderma Spp.* -Cellulase genes- SSF -SmF-CMCase-FPase- β-glucosidase-*CBH2-BGL1.*

INTRODUCTION

Bioethanol from agricultural waste is an attractive way to turn waste into added value that will solve the problem of feed competition and waste management. Napier grass is a highly productive and effective lignocellulosic biomass (Mueansichai et al., 2022). The cellulolytic activity of fungal strains determined by their ability to grow and form clear zones around colonies on Mandel's agar medium (Mandel's mineral solution (MS) supplemented with trace elements, 2% agar and 0.5% w/v low viscosity CMC (Teather & Wood, 1982). For cellulase activity assays, there is always a gap between initial cellulase activity assays and final hydrolysis measurement. To be most meaningful, individual cellulase component assays must be based on a reliable estimation of the amount of individual enzyme component present in the assay. This information permits the calculation of specific activity, i.e., bonds broken per milligram enzyme per unit time. It was showed that the total cellulase system consists of endoglucanases, exoglucanases, and β -D-glucosidases, all of which hydrolyze crystalline cellulose synergically (Liu *et al.*, 2011).

Solid-state fermentation is one of the easiest and cheapest methods for producing microbial bioactive compounds. Trichoderma harzianum has long been recognized as one of the potential fungi for this purpose. Trichoderma spp. was isolated from banana rhizosphere using the soil dilution method and later screened for their ability to produce cellulases using filter paper activity (FPase) and the (CMCase) test. Trichoderma sp. was also subjected to one factor change at a time to determine the effects of different parameters on cellulase production. It was observed that T. harzianum showed the ability to produce higher cellulase activity when wheat straw was used as the substrate. The results showed that 38.5 U/g of cellulase was produced with the use of wheat straw coupled with an incubation temperature of 28 °C and moisture content of 60%. T. harzianum showed solid-state fermentation, with the possibility of its application to industry (Heng & Hamzah, 2022).

CMCase is an amorphous cellulose is used to study CMCase (endoglucanase), while Aciel, an example of crystalline cellulose is used to study FPase (exoglucanases). Isolates produced higher CMCase than FPase even when CMC was used as the only carbon source. Therefore, it could be inferred that the cellulase secreted by methods is an endoglucanase with lesser exoglucanases activities(Kazeem et al., 2021). Lower CMCase activity compared to FPase on the same inducer substrate is a common occurrence in numerous studies. This is because CMC is an amorphous form of cellulose that is easier to digest than filter paper (Oni et al., 2020). The filamentous fungi can grow on solid substrate under solid-state fermentation (SSF) and submerged fermentation (SmF). SSF is regarded as the growth of fungi on solid substrates in the absence of free water. The solid substrates not only supply as nutrients to the fungi but also serve as anchorage for the fungi offers (Gupta, 2017). Additionally, it environments for growth that are comparable to those in which the fungi naturally grow. Moreover, better contact between microorganisms and their substrates can be achieved using SSF, which results in better growth and higher bio production. Industrially important enzymes have traditionally been obtained from submerged fermentation (SmF) because of the ease of handling and greater control of environmental factors such as temperature and pH. However, solid state fermentation (SSF) technique can improve the yield and reduces the cost of enzyme production. There are several reports describing use of agro-industrial residues for the production of cellulose such as wheat straw, wheat bran and rice straw as substrates (Alegre et al., 2009; Kshirsagar et al., 2020; Patel et al., 2019; Yadav et al., 2018) The other advantages of SSF include superior productivity, simple technique; low capital investment, low energy requirement and less water output (Zeng & Chen, 2009; Shad et al., 2023).

The main objectives of this study is to detect the correlation between cellulase activities quantitatively and qualitatively, and screening of (CBH2, BGL1) genes for 26 *Trichoderma* isolates.

MATERIAL AND METHODS

Strains

Twenty-six Trichoderma *spp.* strains obtained from Department of Genetics, Faculty of Agriculture, Menoufia University, Shibeen EL-Kom, Egypt.(El-Sobky *et al.*, 2019)

Qualitative screening (Total cellulases)

Dye staining of carboxy methyl cellulose agar (CMC agar) was autoclaved, dispensed into Petri dishes, allowed to solidify, and inoculated with 5×10^5 spores of *Trichoderma* strains and incubated at 28°C. After 5 days, plates were flooded with 1% aqueous Congo red and allowed to be stained for 15 minutes. The stain was washed off from the agar surface with distilled water and the plates were flooded with 1 M NaCl to destining for 15 minutes. The diameter of the clear zone was measured and recorded (Pointing, 1999).

Walseth cellulose plate-clearing assay this method of (Bose, 1963) with some modifications was carried out to screen for high cellulaseproducing isolates. The basic medium consisted of Mandel's mineral solution MS supplemented with 2% agar. Phosphoric acid-swollen cellulose (PASC) (Walseth cellulose) (Rautela & Cowling, 1966) ,was used as the sole source of carbon at a concentration of 1%. The plates were seeded with 5×10^5 spores in 20µl and incubated at 28°C for 48 hours. Then the diameter of the radial growth was measured and recorded.

Quantitative determination

(CMC) was used as a sole carbon source in submerged fermentation SmF. The measurements were performed after 2,3,4,6 and 7 days of incubation from the filtrate of the inoculated flasks for in Mandel's Media with CMC 0.5% as a substrate of Cellulase. Filter paper Assay (FPase) Total cellulase activity was measured by the filter paper (FPase) method, with Whatman No. 1 filter paper strips (1 cm×6 cm) used as substrate (Ghose, 1987). Endoglucanase (CMCase) CMC (2% w/v) in 50 mM citrate buffer pH.4.8 (freshly made) was used as substrate solutions for Endoglucanase assays. 0.5 mL of the culture filtrate was transferred into test tube with a volume of at least 10 mL. The culture filtrate

solution and substrate solution were equilibrated at 50°. (Miller, 1959). ß-glucosidase activity assay was determined according to (Ghose, 1987)as follows: Cellobiose (15 mM) in 50 mM citrate buffer pH.4.8 (freshly made) was used as substrate solutions for β -Glucosidase assays .

Solid state fermentation

The solid-state culture was kept in 250-mL flasks; the flasks were closed with a cotton covered plug. Each flask contained 3 g of straw and 12 mL Mandel's basal salt solution (MS). The flasks were autoclaved at 121°C for 30 min. Each flask was then inoculated with 1 mL conidial suspension to produce a final concentration of 1×10^7 conidia. The flasks were incubated at 30 °C with a relative humidity of 70% for 10 days.

Detection of Cellulase genes of *Trichoderma* species

Trichoderma isolates were cultured on PDA broth at 28 °C for 5 days, after which total genomic DNA for each strain was extracted, using method reported by (Elsobky *et al.*, 2019). Amplification of Chb gene was achieved according to (Wang & Gao, 1999) and the Bgl gene was achieved according to (González *et al.*, 1992) primer sequences, amplification conditions, and amplicon sizes listed in

Data analysis

All experiments that were conducted in this study were implemented in accordance with the Completely Randomized Design (CRD). The determination of significant differences was calculated (Duncan Test). Statistical tests were performed using a statistical analysis by computer program system developed by IBM Corp. in 2017, specifically IBM SPSS Statistics for Windows, Version 25.0, denoting their significance at P < 0.05.

RESULTS AND DISCUSSION

Qualitative measurement of cellulase production

Trichoderma isolates were identified for their endoglucanase activities on plate by using assay that appeared when using (CMC) and Congo red dye. Based on measurements of clear zone diameter, six isolates gave clear zones of cellulase activity having diameter significantly larger than other isolates namely, MNF-MAS-*Tricho*1, MNF-MAS-*Tricho*6, MNF-MAS-*Tricho*23, and ASI-MAS-*Tricho*26 Figure (1).



a)



b)

Figure (1): a) Endoglucanase activities on plate by using (CMC) staining by Congo red dye.
b) Radial growth of Trichoderma Isolates in Mandel's Media containing PASC Phosphoric acid-swollen cellulose 1%.

Data presented in Table (1) showed that (CMC) was a substrate for endoglucanase and so can be used as a test for endoglucanase activity and the 1% PASC media full plate radial growth showed in isolates MNF-MAS-*Tricho2*, 3, 5, 6, 12, 15, 18, 21, and MNF-MAS-*Tricho23* after 48 hrs from incubation and the detection of the sporulation type showed green spores in 7 isolates

Table (1). Various studies based on method of Congo red to screen the different *Trichoderma* cellulolytic isolates (Castrillo *et al.*, 2017; Suirta *et al.*, 2021) Avicel as a" PASC "has been used in our study to be measuring exo-glucanase activities that cleave the accessible ends of cellulose modules to liberate glucose and cellobiose (Fahmi *et al.*, 2016; Sharrock, 1988).

 Table (1): Qualitative screening of Trichoderma spp. isolates using (CMC), and micro-crystalline cellulose as substrates.

		lucanase	Sporulation type in 1%
<i>Trichoderma</i> isolate	PASC	ameter in cm) * CMC	PASC Media
MNF-MAS-Tricho1	0.30 a	5.83 ^{jk}	Green spores*
MNF-MAS- Tricho 2	9.00 ^h	2.97 ^{de}	No spores
MNF-MAS- Tricho 3	9.00 ^h	2.83 ^d	No spores
MNF-MAS- Tricho 4	7.00 ^g	4.20 ^h	Green spores*
MNF-MAS- Tricho 5	9.00 ^h	0.17ª	Green spores*
MNF-MAS- Tricho 6	9.00 ^h	5.57 ^{jk}	Light green spores *
MNF-MAS- Tricho 7	5.00 ^e	3.73 ^{fg}	No spores
GIZ-MAS- Tricho 8	3.00 ^d	4.20 ^h	No spores
MNF-MAS- Tricho 9	6.20 ^f	4.00 ^{gh}	No spores
MNF-MAS- Tricho 10	7.10 ^g	3.40 ^{ef}	No spores
MNF-MAS- Tricho 11	5.20 ^e	4.00 ^{gh}	Green spores**
MNF-MAS- Tricho 12	9.00 ^h	4.93 ⁱ	Green spores**
MNF-MAS- Tricho 13	7.13 ^g	1.17 ^b	No spores
MNF-MAS- Tricho 14	5.00 ^e	1.97 °	No spores
MNF-MAS- Tricho 15	9.00 ^h	1.43 ^b	Green spores**
MNF-MAS- Tricho 16	0.10 ^a	0.17 ª	No spores
MNF-MAS- Tricho 17	1.10 ^b	0.27 ª	No spores
MNF-MAS- Tricho 18	9.00 ^h	4.43 ^h	No spores
MNF-MAS- Tricho 19	6.13 ^f	5.00 ⁱ	No spores
MNF-MAS- Tricho 20	0.27 ^a	3.00 ^{de}	No spores
MNF-MAS- Tricho 21	9.00 ^h	5.43 ^j	No spores
MNF-MAS- Tricho 22	0.17 ^a	2.10 °	No spores
MNF-MAS- Tricho 23	9.00 ^h	5.93 ^k	Light green spores **
GIZ-MAS-2 Tricho 24	2.00°	4.33 h	No spores
AST-MAS- Tricho 25	0.00ª	4.43 ^h	No spores
AST-MAS- Tricho 26	6.10 ^f	5.93 ^k	Green spores **

*Within columns, values with a common letter do not differ significantly (P<0.05), according to Duncan_{a,b,c}

Correlation between Qualitative and Quantitative Cellulase Enzymes activities in some *Trichoderma spp*.

	Jaclata Cada	Enzymes activities (IU mL ⁻¹)				
	Isolate Code	2 days	3 days	4 days	6 days	7 days
1	MNF-MAS-Tricho1	0.28 ^f	0.30 e	0.31 ^{ef}	0.34 ^{gh}	0.43 efg
2	MNF-MAS- Tricho 2	0.18 ^a	0.31 ef	0.37 ⁱ	0.45 ^j	0.49 ^g
3	MNF-MAS- Tricho 3	0.26 ^{ef}	0.28 °	0.32 ^{fg}	0.27 ^{cd}	0.47 ^g
4	MNF-MAS- Tricho 4	0.25 ^{def}	0.27 ^{bc}	0.32 ^{fg}	0.27 ^{cd}	0.43 efg
5	MNF-MAS- Tricho 5	0.24 ^{cde}	0.26 ^{ab}	0.28 ^{bc}	0.24 ^{ab}	0.43 efg
6	MNF-MAS- Tricho 6	0.24 ^{cdef}	0.38 ^h	0.40 ^j	0.27 ^{cd}	0.40 ^{def}
7	MNF-MAS- Tricho 7	0.22 ^{bcd}	0.27 ^{bc}	0.41 ^j	0.30 e	0.44 efg
8	GIZ-MAS- Tricho 8	0.21 abc	0.25 ^a	0.25 ª	0.26 ^{cd}	0.26 ^a
9	MNF-MAS- Tricho 9	0.25 def	0.27 ^{bc}	0.34 ^h	0.25 bc	0.38 cde
10	MNF-MAS- Tricho 10	0.24 ^{cde}	0.29 ^{de}	0.28 ^b	0.35 ^h	0.38 cde
11	MNF-MAS- Tricho 11	0.19 ^{ab}	0.24 ^a	0.25 ª	0.26 ^{cd}	0.26 ^a
12	MNF-MAS- Tricho 12	0.25 ^{def}	0.27 ^{bc}	0.28 ^b	0.33 ^{fg}	0.35 ^{cd}
13	MNF-MAS- Tricho 13	0.23 bcde	0.27 °	0.30 ^{def}	0.23 ^a	0.45 ^{fg}
14	MNF-MAS- Tricho 14	0.19 ^{ab}	0.24 ^a	0.25 a	0.26 ^{cd}	0.26 ^a
15	MNF-MAS- Tricho 15	0.31 ^f	0.31 ef	0.32 ^{fg}	0.27 ^{cd}	0.47 ^g
16	MNF-MAS- Tricho 16	0.21 abc	0.24 ^a	0.25 ^a	0.26 ^{cd}	0.26 ^a
17	MNF-MAS- Tricho 17	0.35 ^g	0.25 ^{ab}	0.30 ^{cde}	0.28 ^d	0.45 ^{fg}
18	MNF-MAS- Tricho 18	0.27 ^{ef}	0.33 ^g	0.33 ^{gh}	0.42 ⁱ	0.34 ^{cd}
19	MNF-MAS- Tricho 19	0.25 def	0.31 ef	0.32 ^{fg}	0.27 ^{cd}	0.33 abc
20	MNF-MAS- Tricho 20	0.25 def	0.28 ^{cd}	0.33 ^{gh}	0.30 e	0.35 ^{cd}
21	MNF-MAS- Tricho 21	0.26 ^{ef}	0.28 ^{cd}	0.29 bcd	0.33 ^{gh}	0.28 ^{ab}
22	MNF-MAS- Tricho 22	0.24 ^{cdef}	0.27 ^{bc}	0.27 ^b	0.26 ^{cd}	0.34 ^{cd}
23	MNF-MAS- Tricho 23	0.25 ^{def}	0.36 ^h	0.36 ⁱ	0.31 ^e	0.34 bcd
24	GIZ-MAS-2 Tricho 24	0.25 def	0.32 ^{fg}	0.30 ^{cde}	0.31 ^{ef}	0.32 ^{abc}
25	AST-MAS- Tricho 25	0.21 abc	0.24 ^a	0.25 ª	0.25 ^{bc}	0.34 bcd
26	AST-MAS- Tricho 26	0.20 ^{ab}	0.24 ^a	0.25 ^a	0.26 ^{cd}	0.40 ^{def}

Table (2): Filter paper enzyme activity FPase in Mandel's Media with CMC 0.5% .

*Within columns, values with a common letter do not differ significantly (P<0.05), according to Duncan_{a,b,c}

Quantitative measurement of cellulase from *Trichoderma*.

All isolates were screened for enzyme activity (filter paper activity (FPase) assay, CMCase for endo- β - 1,4-gluconase and cellobiase assay for β -glucosidase). Therefore, submerged fermentation experiment was carried out by 26 *Trichoderma* isolates.

Cellulase production in submerged fermentation (SmF) cultures.

The measurements were taken after 2,3,4,6 and 7 days of incubation from the filtrate of the inoculated flasks for in Mandel's Media with CMC 0.5% as a substrate of Cellulase. Results represented in the Table (2,3 and 4) showed every enzyme separated and the kinetic study of the best isolates in enzyme production.

Moreover, data presented in Table (2) showed cellulases enzyme activity (measured as FPase after different incubation periods), free sugar produced in Submerged fermentation (SmF) cultures for all isolates. Generally, all isolates indicated outstanding FPase activities. Four isolates demonstrated highly significant FPase activities after 7 days of incubation namely, MNF-MAS-*Tricho*2, MNF-MAS-*Tricho*3 MNF-MAS-*Tricho*15 and MNF-MAS-*Tricho*17(0.49, 0.47, 0.47 and 0.45 IU mL⁻¹, respectively). Whereas

isolates GIZ-MAS-*Tricho*8, MNF-MAS-*Tricho*11, MNF-MAS-*Tricho*14, MNF-MAS-*Tricho*16 and MNF-MAS-*Tricho*21 showed the lowest FPase activity (0.26 IU mL⁻¹) were in average values. MNF-MAS-*Tricho*2 showed the highest level of FPase after 4, 6, 7 days of incubation (0.37,0.45 and 0.49 IU mL⁻¹, respectively). MNF-MAS-*Tricho*15 showed the high level of FPase in two, seven days of incubation (0.31and 0.47 IU mL⁻¹, respectively).

Moreover, MNF-MAS-*Tricho*17 showed the high level of FPase in 2, 7days of incubation they came to be (0.35 and 0.45 IU mL⁻¹, respectively). MNF-MAS-*Tricho*7 showed a high level of FPase activity after four days of incubation (0.41 IU mL⁻¹) but in the remaining days of incubation didn't show any high levels of FPase. This Table (2) showed that the high significant values of FPase enzyme activity came after 6 days of incubation.

	Lash-ta Cash-		Enzymes activities (IU mL ⁻¹)				
	Isolate Code	2 days	3 days	4 days	6 days	7 days	
1	MNF-MAS-Tricho1	0.43 ^{cdef}	0.43 abc	0.48 ^{ab}	0.52 ^{ab}	0.59 cdef	
2	MNF-MAS- Tricho 2	0.37 ^{ab}	0.43 abc	0.53 bcde	0.53 ^{abc}	0.61 efgh	
3	MNF-MAS- Tricho 3	0.51 ^{hi}	0.53 ^e	0.51 bcd	0.54 ^{bc}	0.65 ^{ghij}	
4	MNF-MAS- Tricho 4	0.45 ^{defg}	0.43 abc	0.51 bcd	0.64 ^f	0.69 ^j	
5	MNF-MAS- Tricho 5	0.38 abc	0.44 ^{abcd}	0.56 defgh	0.60 ^{ef}	0.75 ^k	
6	MNF-MAS- Tricho 6	0.38 abc	0.45 abcd	0.59 ^{fghi}	0.64 ^f	0.801	
7	MNF-MAS- Tricho 7	0.51 ^{hi}	0.46 bcd	0.62 ⁱ	0.62 ef	0.67 ^{ij}	
8	GIZ-MAS- Tricho 8	0.41 abcd	0.44 ^{abcd}	0.45 ª	0.49 ^a	0.51 ^a	
9	MNF-MAS- Tricho 9	0.47 ^{fghi}	0.44 ^{abcd}	0.60 ^{hi}	0.60 ef	0.64 ^{ghi}	
10	MNF-MAS- Tricho 10	0.51 ^{hi}	0.44 ^{abcd}	0.58 ^{fghi}	0.54 ^{bc}	0.60 defg	
11	MNF-MAS- Tricho 11	0.41 abcd	0.44 ^{abcd}	0.45 ª	0.49 ^a	0.51 ^a	
12	MNF-MAS- Tricho 12	0.49 ^{hi}	0.43 abc	0.57 efghi	0.55 bcd	0.55 abed	
13	MNF-MAS- Tricho 13	0.36 ^a	0.41 ^a	0.58 efghi	0.55 bcd	0.62 efgh	
14	MNF-MAS- Tricho 14	0.41 abcd	0.44 ^{abcd}	0.45 ^a	0.49 ^a	0.51 ^a	
15	MNF-MAS- Tricho 15	0.43 ^{cdef}	0.45 abcd	0.58 efghi	0.59 ^{de}	0.63 fghi	
16	MNF-MAS- Tricho 16	0.41 ^{abcd}	0.44 abcd	0.45 ª	0.49 ^a	0.51 ^a	
17	MNF-MAS- Tricho 17	0.52 ⁱ	0.42 ^{ab}	0.53 ^{bcde}	0.53 abc	0.58 ^{bcde}	
18	MNF-MAS- Tricho 18	0.47 ^{fghi}	0.46 bcd	0.50 abc	0.53 ^{ab}	0.59 cdef	
19	MNF-MAS- Tricho 19	0.46 ^{efgh}	0.48 ^d	0.51 ^{bcd}	0.52 ^{ab}	0.60^{defg}	
20	MNF-MAS- Tricho 20	0.47 ^{fghi}	0.46 ^{cd}	0.54 ^{cdef}	0.53 abc	0.55 abcd	
21	MNF-MAS- Tricho 21	0.51 ^{hi}	0.61 ^f	0.57 efghi	0.55 bcd	0.55 abc	
22	MNF-MAS- Tricho 22	0.41 bcde	0.68 ^g	0.70 ^j	0.74 ^g	0.90 ^m	
23	MNF-MAS- Tricho 23	0.41 bcde	0.44 ^{abcd}	$0.54^{\text{ cdefg}}$	0.53 abc	0.53 ^a	
24	GIZ-MAS-2 Tricho 24	0.44 ^{def}	0.48 ^d	0.50 abc	0.55 bcd	0.59 ^{cdef}	
25	AST-MAS- Tricho 25	0.49 ^{ghi}	0.44 ^{abcd}	0.59 ^{ghi}	0.58 ^{de}	0.61 efgh	
26	AST-MAS- Tricho 26	0.44 ^{def}	0.44 ^{abcd}	0.49 abc	0.52 ^{ab}	0.54 ^{ab}	

Table (3) CMCase activity in Mandel's Media with CMC 0.5%.

*Within columns, values with a common letter do not differ significantly (P<0.05), according to Duncanabe

Isolate Code Enzymes activities (IU mL ⁻¹)						
	Isolate Coue	2 days	3 days	4 days	6 days	7 days
1	MNF-MAS-Tricho1	0.34 ^a	0.30 ª	0.65 ^{hi}	0.61 ^{efg}	0.59 abcd
2	MNF-MAS- Tricho 2	0.61 ⁱ	0.62 ^d	0.65 ^{hi}	0.66 ^{hi}	0.75 ^{hi}
3	MNF-MAS- Tricho 3	0.53 ^{fg}	0.45 ^b	0.51 ^{ab}	0.59 ^{cdef}	0.70^{fgh}
4	MNF-MAS- Tricho 4	0.58 ^{hi}	0.46 ^b	0.52 abc	0.53 ª	0.73 ^{ghi}
5	MNF-MAS- Tricho 5	0.48 ^{cde}	0.47 ^b	0.55 bcd	0.62 ^{fgh}	0.72 ^{ghi}
6	MNF-MAS- Tricho 6	0.55 ^{gh}	0.60 ^d	0.59 efg	0.59 ^{bcdef}	0.67 ^{efg}
7	MNF-MAS- Tricho 7	0.45 ^{cd}	0.49 ^b	0.51 ^{ab}	0.53 ^a	0.53 ^a
8	GIZ-MAS- Tricho 8	0.47 ^{cde}	0.48 ^b	0.59 efg	0.60 ^{def}	0.65 ^{def}
9	MNF-MAS- Tricho 9	0.48 ^{cde}	0.47 ^b	0.55 ^{cde}	0.62 ^{fg}	0.76 ⁱ
10	MNF-MAS- Tricho 10	0.45 ^{cd}	0.49 ^b	0.51 ^{ab}	0.53 ^a	0.53 ^a
11	MNF-MAS- Tricho 11	0.48 ^{de}	0.47 ^b	0.53 ^{abc}	0.55 ^{abc}	0.63 ^{cde}
12	MNF-MAS- Tricho 12	0.46 cde	0.47 ^b	0.55 bcd	0.57 abcde	0.61 bcde
13	MNF-MAS- Tricho 13	0.45 ^{cd}	0.49 ^b	0.51 ^{ab}	0.53 ^a	0.53 ^a
14	MNF-MAS- Tricho 14	$0.54^{\text{ gh}}$	0.54 °	0.53 ^{abc}	0.60 ^{def}	0.72 ^{ghi}
15	MNF-MAS- Tricho 15	0.45 ^{cd}	0.49 ^b	0.51 ^{ab}	0.53 ^a	0.53 ^a
16	MNF-MAS- Tricho 16	0.54 ^{gh}	0.47 ^b	0.62 ^{gh}	0.64 ^{ghi}	0.62 ^{cde}
17	MNF-MAS- Tricho 17	0.58 ^{hi}	0.62 ^d	0.73 ^j	0.77 ^{ij}	0.75 ^{hi}
18	MNF-MAS- Tricho 18	0.56 ^{gh}	0.60 ^d	0.60 ^{fg}	0.62 ^{fgh}	0.70 ^{fghi}
19	MNF-MAS- Tricho 19	0.50 ^{ef}	0.48 ^b	0.58 def	0.60 ^{defg}	0.90 ^j
20	MNF-MAS- Tricho 20	0.40 ^b	0.49 ^b	0.80 k	0.67 ⁱ	0.58 abc
21	MNF-MAS- Tricho 21	0.34 ^a	0.54 °	0.71 ^j	0.62 ^{fgh}	0.55 ^{ab}
22	MNF-MAS- Tricho 22	0.45 ^{cd}	0.62 ^d	0.62 ^{gh}	0.62 ^{fgh}	0.61 bcde
23	MNF-MAS- Tricho 23	0.31 ^a	0.48 ^b	0.66 ⁱ	0.62 ^{fgh}	0.58 abc
24	GIZ-MAS-2 Tricho 24	0.44 ^{bc}	0.48 ^b	0.50 ^a	0.55 ^{ab}	0.59 abc
25	AST-MAS- Tricho 25	0.46 ^{cde}	0.48 ^b	0.52 ^{abc}	0.56 abcd	0.61 bcde
26	AST-MAS- Tricho 26	0.47 ^{cde}	0.47 ^b	0.53 ^{abc}	0.55 ^{ab}	0.60 ^{abcd}

Table (4): β-Glucosidase in Mandel's Media with CMC 0.5%.

*Within columns, values with a common letter do not differ significantly (P<0.05), according to Duncana,b,c

Data presented in Table (3) showed cellulases enzyme activity (measured as CMCase after different incubation periods), Generally, all isolates indicated outstanding endoglucanase activity. While MNF-MAS-Tricho22 isolate showed the highest significant CMCase activity (0.90 IU mL⁻¹) after seven days of incubation. Furthermore, this isolate showed high level of CMCase after 3,4 and 6 days of incubation (0.68, 0.70 and 0.74 IU mL¹, respectively). Those results clearly showed that isolates MNF-MAS-Tricho6 and MNF-MAS-Tricho5. The lowest level of CMCase appeared after incubation seven isolates GIZ-MAS-Tricho8, MNF-MAS-Tricho11, MNF-MAS-Tricho14, MNF-MAS-Tricho16 MNF-MAS-Tricho23 and AST-MAS-

*Tricho*26 all of them come to be lower than the above tested isolates (0.55IU mL⁻¹). MNF-MAS-*Tricho*17 showed higher enzyme activity after two days of growth as it came to be 0.52 IUmL⁻¹. In fact, this isolate showed steady of enzyme activity as they came to be 0.52, 0.42, 0.53, 0.53 and 0.58 IU mL⁻¹ respectively in different days of the measurement.

Table (4) showed the cellulases enzyme activities (β -glucosidase after four different incubation periods), MNF-MAS-*Tricho*19 was the highest (0.90 IU mL⁻¹) while MNF-MAS-*Tricho*7, MNF-MAS-*Tricho*10, MNF-MAS-*Tricho*13, MNF-MAS-*Tricho*15, MNF-MAS-*Tricho*23 and GIZ-MAS-*Tricho*24 were the

lowest (0.5 IU mL⁻¹) after 7 days of incubation. MNF-MSH-*Trich*2 showed the highest value for β -glucosidase enzyme activity in the culture filtrate at the beginning of the experiment after two days and three days their values come to be (0.61 and 0.62 IU mL⁻¹, respectively), While most isolates showed low level of the enzyme activities, at the first sample after two days and the end of experiment ,whereas it showed high significant values of β -glucosidase enzyme activity after 7 days of growth .

Successful enzymatic hydrolysis of cellulose necessitates the combined action of exoglucanases. endoglucanases, and betaglucosidase (Zhang et al., 2018). To evaluate the activities of FPase, CMCase, and β-glucosidase; Qualitative and quantitative assays were conducted on minimal media supplemented with sugar cane bagasse and rice straw as the sole carbon sources. The results obtained from both types of assays were highly consistent with those reported by (Sazci et al., 1986; Castrillo et al., 2021). They affirmed the congruence between qualitative and quantitative methods and have recommended the use of the clearing zone assay as a fast, cost-effective, and sensitive test for screening many isolates. In a study conducted by (Marecik et al., 2018), 123 strains of Trichoderma were tested for their ability to degrade cellulose. Approximately 30 strains exhibited significantly higher levels of cellulase activities compared to the reference strain. The variation in cellulolytic activities among Trichoderma isolates may be attributed to differences isolates in genetic content, origin, and the number of cellulase enzymes that present in tested fungus.

Our obtained data agreed nicely with what reported by Nathan *et al.*, (2014) they demonstrated that the optimum enzyme recovery period was identified between 5th to 9th days of incubation.

Zahra *et al.*, (2020) demonstrated that the Cellulase Production by *Trichoderma viride* in Submerged Fermentation SmF represents as cheaper process and the good alternative for industrial applications. These findings were consistent with the result that most of the isolates belonged to *T. longibarchiatum* and *T. harzianum*. These species have been adopted in various industries because of their ability to secrete large amounts of protein and metabolites (Gupta *et al.*, 2014) Nutrient supplies have been reported to be key elements in the synthesis of cellulase and xylanase carbon source is an essential nutrient affecting enzyme production. Moreover, the pure CMC using as sole carbon source represents a suitable substrate for quantitative cellulase activity measurements in SmF (Dewiyanti *et al.*, 2022).

Avicel has been used for measuring exoglucanase activities that cleave the accessible ends of cellulose modules to liberate glucose and cellobiose (Sharrock, 1988).

Figure (2) Showed mean cellulase produce from tested *Trichoderma* isolates in Solid state fermentation (SSF) with the wheat and rice straw after different incubation days. The soft mycelium of *Trichoderma* showed for the first time after three days and the sporulation detected with the green color observed on the straw after five days. The hydrolysis of the rice and wheat straw (weight loss and glucose free sugar) is shown in Table (5) and the enzyme production (FPase, CMCase and β -glucosidase) showed in Table (6).

Table (5) showed the total weight loss and free glucose sugar produced in (SSF) rice and wheat straw by Trichoderma isolates. There were high variations in losing weight from (4% to 40%) in rice straw after 10 days of incubation and showed differences in glucose concentration levels are ranging from (1.3: 3.14 mg/ml). Isolate MNF-MAS-Tricho18 shows a high level on both substrate (34.56% and 3.14 mg/ml). Moreover, there were high variations weight loss in wheat straw from (6% to 20%) after 10 days of incubation that showed lower than weight loss in rice straw (4% to 39%) in the same incubation condition. Glucose concentrations are ranging from (0.24 to 5mg/ml). Isolate MNF-MAS-Tricho3 showed high level in both (4.32mg/ml and 19.6%).

Correlation between Qualitative and Quantitative Cellulase Enzymes activities in some *Trichoderma spp*.

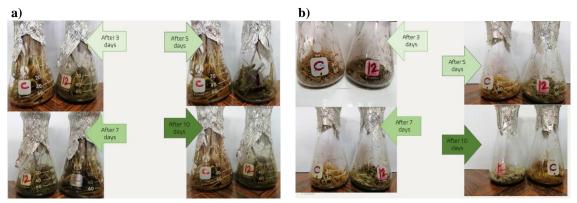


Figure (2): Solid state fermentation (SSF) of some tested *Trichoderma* strain MNF-MAS- *Tricho* 12 a) Rice straw b) Wheat straw.

Table (5): Total weight loss and	glucose free sugar	produced in	(SSF)	rice and	wheat	straw by
Trichoderma strains af	iter 10 days.					

		Rice str	aw	Wheat	straw
	Isolates	Weight loss %	Glucose mg/ml	Weight loss %	Glucose mg/ml
1	MNF-MAS-Tricho1	31.72% ^{jk}	2.01 ^{ghi}	16.62% ^{ij}	7.47 ^k
2	MNF-MAS- Tricho 2	29.29% ^{hi}	2.13 ⁱ	19.88% ¹	0.90 ^d
3	MNF-MAS- Tricho 3	24.56% ^g	1.45 ^{abcd}	19.60% ^{kl}	4.32 ⁱ
4	MNF-MAS- Tricho 4	33.79% ^{kl}	1.86 ^{fg}	13.53% ^{fg}	1.45 ^e
5	MNF-MAS- Tricho 5	21.83% ^f	1.76 ^f	18.02% ^{jk}	0.44 ^{ab}
6	MNF-MAS- Tricho 6	4.38% ^a	2.07 ^{hi}	14.09% fgh	4.30 ⁱ
7	MNF-MAS- Tricho 7	11.18% ^c	1.94 ^{gh}	10.72% ^{cd}	0.24 ^a
8	GIZ-MAS- Tricho 8	8.28% ^b	1.36 ^{ab}	10.16% bc	0.24 ^a
9	MNF-MAS- Tricho 9	12.90% ^c	1.41 abcd	8.70% ^b	3.63 ^h
10	MNF-MAS- Tricho 10	39.59% ^m	1.56 ^{cd}	12.97% ^{ef}	0.77 ^{cd}
11	MNF-MAS- Tricho 11	15.38% ^d	1.39 abc	10.16% ^{bc}	0.30 ^a
12	MNF-MAS- Tricho 12	30.77% ^{ij}	2.54 ^{ik}	9.38% ^{bc}	1.81 ^f
13	MNF-MAS- Tricho 13	15.27% ^d	1.44 ^{abcd}	5.67% ^a	0.26 ^a
14	MNF-MAS- Tricho 14	7.69% ^b	1.37 ^{ab}	10.16% ^{bc}	0.30 ^a
15	MNF-MAS- Tricho 15	17.75% ^e	2.05 ^{hi}	10.56% ^{bc}	4.90 ^j
16	MNF-MAS- Tricho 16	34.32% ¹	1.30 ^a	9.60% ^{bc}	0.27 ^a
17	MNF-MAS- Tricho 17	6.69% ^b	2.55 ^{ik}	13.25% ^f	0.68 ^{bcd}
18	MNF-MAS- Tricho 18	34.56% ¹	3.14 ¹	17.91% ^{jk}	0.62 ^{bc}
19	MNF-MAS- Tricho 19	12.60% °	1.51 bcd	10.50% ^{bc}	1.41 ^e
20	MNF-MAS- Tricho 20	26.98% ^h	1.59 ^d	15.78% ^{hi}	1.86 ^f
21	MNF-MAS- Tricho 21	27.75% ^h	2.71 ^k	18.59% ^{kl}	2.17 ^g
22	MNF-MAS- Tricho 22	18.34% ^e	2.15 ⁱ	18.02% ^{jk}	2.35 ^g
23	MNF-MAS- Tricho 23	34.32% ¹	2.16 ⁱ	13.25% ^f	4.25 ⁱ
24	GIZ-MAS-2 Tricho 24	28.93% ^{hi}	2.40 ^j	11.29% cde	2.22 ^g
25	AST-MAS- Tricho 25	26.98% ^h	1.36 ^{ab}	11.29% cde	0.24 ^a
26	AST-MAS- Tricho 26	7.69% ^b	2.71 ^k	12.41% ^{def}	2.17 ^g

*Within columns, values with a common letter do not differ significantly (P<0.05), according to Duncan_{a,b,c}

		С	ellulase enzymes	activity activiti	es (IU g ⁻¹)	
Isolates		Rice straw Wheat straw			1	
	FPase	CMCase	β-Glucosidase	FPase	CMCase	β-Glucosidase
MNF-MAS-Tricho1	0.085 ^{gh}	0.120 ab	0.076ª	0.063 ^{ef}	0.080 ^{ab}	0.078 ^{abc}
MNF-MAS- Tricho 2	0.114 ^j	0.156 ^{hij}	0.154 ^e	0.076 ⁱ	0.104 ^{hij}	0.103 ^d
MNF-MAS- Tricho 3	0.068 abcde	0.127 abcd	0.112 bcd	0.065 ^{fgh}	0.084 abcd	0.075 ^{abc}
MNF-MAS- Tricho 4	0.069 ^{bcde}	0.127 abcd	0.114 bcd	0.046 bc	0.084 abcd	0.076 abc
MNF-MAS- Tricho 5	0.062 ^{ab}	0.139 ^{cdefg}	0.116 bcd	0.041 ^{ab}	0.093 ^{cdefg}	0.078 abc
MNF-MAS- Tricho 6	0.102 ⁱ	0.149 ^{fghi}	0.149 ^e	0.068 fgh	0.099 fghi	0.099 ^d
MNF-MAS- Tricho 7	0.075 ^{ef}	0.154 ^{ghi}	0.106 ^{bc}	0.050 ^{cd}	0.103 ^{ghi}	0.071 ^{ab}
GIZ-MAS- Tricho 8	0.067 abed	0.112ª	0.120 ^{cd}	0.044 abc	0.075 ^a	0.080 bc
MNF-MAS- Tricho 9	0.064 ^{abc}	0.149 ^{fghi}	0.117 bcd	0.043 ^{ab}	0.099 fghi	0.078 ^{abc}
MNF-MAS- Tricho 10	0.088 ^h	0.146 efghi	0.114 bcd	0.059 ^e	0.097 ^{efgh}	0.076 abc
MNF-MAS- Tricho 11	0.067 abed	0.112 a	0.117 bcd	0.044 abc	0.075 ^a	0.078 ^{abc}
MNF-MAS- Tricho 12	0.103 ⁱ	0.159 ^{ij}	0.147 ^e	0.069 ^{gh}	0.106 hij	0.098 ^d
MNF-MAS- Tricho 13	0.060 ^a	0.143 ^{efgh}	0.106 bc	0.040 ^a	0.096 ^{efgh}	0.071 ^{ab}
MNF-MAS- Tricho 14	0.067 abed	0.112 a	0.110 bcd	0.044 abc	0.075 ^a	0.073 ^{abc}
MNF-MAS- Tricho 15	0.067 abed	0.143 efgh	0.123 ^d	0.045 abc	0.096 ^{efgh}	0.082°
MNF-MAS- Tricho 16	0.067 abed	0.112 a	0.108 bcd	0.044 abc	0.075 ^a	0.072 ^{abc}
MNF-MAS- Tricho 17	0.070 ^{cde}	0.131 ^{bcde}	0.108 bcd	0.047 ^{bc}	0.087 ^{bcde}	0.072 ^{abc}
MNF-MAS- Tricho 18	0.105 ⁱ	0.140 ^{cdefg}	0.151 ^e	0.070 ^h	0.094 cdefg	0.101 ^d
MNF-MAS- Tricho 19	0.068 ^{bcde}	0.126 abc	0.111 bcd	0.045 abc	0.084 abc	0.074 ^{abc}
MNF-MAS- Tricho 20	0.074 ^{def}	0.134 ^{bcdef}	0.105 ^b	0.050 ^{cd}	0.089 ^{bcdef}	0.070 ^a
MNF-MAS- Tricho 21	0.084 ^{gh}	0.141 ^{defgh}	0.109 bcd	0.066 fgh	0.094 ^{defgh}	0.073 abc
MNF-MAS- Tricho 22	0.066 abc	0.176 ^k	0.105 ^b	0.064 ^{fg}	0.117 ^k	0.070 ^a
MNF-MAS- Tricho 23	0.103 ⁱ	0.168 ^{jk}	0.153 ^e	0.069 ^{gh}	0.112 ^{jk}	0.103 ^d
GIZ-MAS-2 Tricho 24	0.079 ^{fg}	0.124 ^{ab}	0.110 bcd	0.052 ^d	0.082 ^{ab}	0.073 abc
AST-MAS- Tricho 25	0.064 abc	0.147 ^{fghi}	0.116 bcd	0.043 ^{ab}	0.098 ^{fghi}	0.077 ^{abc}
AST-MAS- Tricho 26	0.067 abcd	0.123 ab	0.119 bcd	0.044 abc	0.082 ^{ab}	0.079 abc

Table (6): Total Cellulase enzymes activity (IU g-1) (FPase, CMCase and β-glucosidase), in (SSF) Rice
and wheat straw by <i>Trichoderma</i> strains after 10 days.

*Within columns, values with a common letter do not differ significantly (P<0.05), according to Duncana,b,c

Data presented in Table (6) clearly showed the cellulases enzyme activities (measured as FPase, CMCase and β -glucosidase) in SSF (Rice straw) for all isolates. Generally, all isolates indicated outstanding endoglucanase activity, when compared to FPase and β -glucosidase. Five isolates demonstrated highly significant FPase activities namely, MNF-MAS-*Tricho2*, 6,12,18 and 23 (0.114, 0.102, 0.103, 0.105 and 0.103 IU g⁻¹, respectively). MNF-MAS-Tricho 13 showed the lowest FPase activity (0.06 IU g⁻¹). While MNF-MAS-*Tricho2*,12,22 and 23 showed the highest significant CMCase activity (0.156,

0.159, 0.176 and 0.168 IU g⁻¹, respectively) and the lowest were GIZ-MAS-*Tricho* 8,11,14, and 16 (0.112 IU g⁻¹). As for β -glucosidase MNF-MAS-*Tricho* 2,6,12,18 and 23 were the highest in the same level without significant (0.15 IU g⁻¹) while MNF-MAS-*Tricho*1 were the lowest (0.076 IU g⁻¹).

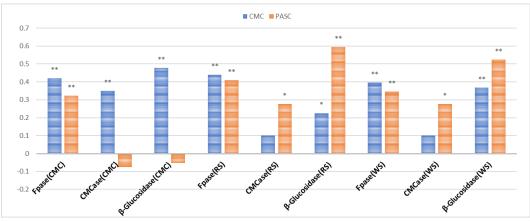
In SSF of wheat straw two isolates demonstrated highly significant FPase activities namely, MNF-MAS-*Tricho*2, and18 (0.076 and 0.070 IU g⁻¹, respectively). MNF-MAS-*Tricho* 13 showed the lowest FPase activity (0.04 IU g⁻¹).

While MNF-MAS-*Tricho* 22 and 23 showed the highest significant CMCase activity (0.112 and 0.117 IU g⁻¹, respectively) and the lowest were GIZ-MAS-*Tricho* 8, MNF-MAS- *Tricho* 11,14, and 16 (0.075 IU g⁻¹). As for β -glucosidase MNF-MAS-*Tricho* 2 and 23 was the highest in the same level without significant (0.103 IU g⁻¹) while MNF-MSH-*Tricho*20 are the lowest (0.070 IU g⁻¹).

It has been revealed that most filamentous fungi are able to grow well under SSF and produce higher levels of extracellular enzymes and also other important metabolites than those under submerged fermentation (SmF). These findings were consistent with the results that most of the isolates belonged to T. longibarchiatum and T. harzianum. These species have been adopted in various industries because of their ability to secrete large amounts of protein and metabolites (Gupta et al., 2014). Moreover, the utilization of rice straw for cellulase production has the potential to mitigate national rice straw waste, while also offering cost-effective advantages to the enzyme industry (Naher et al., 2021). Different methods and different substrates have been used to compare between SSF and SmF for enzyme production; Firstly, Using SmF for enzyme production gave high yield and high quality. Different substrate showed higher enzymatic activity when compared on CMC

substrate than the microcrystalline cellulose (Pirzadah *et al.*, 2014). The results showed that the activities of Filter paper enzyme (FPase), Endoglucanase (CMCase) and β -glucosidase were significantly affected by substrate mixture (Oni *et al.*, 2020)*T. reesei* shows higher enzymatic activity when compared with *T. viride* on CMC substrate than the microcrystalline cellulose. (Pirzadah *et al.*, 2014).

Data in Figure (3) showed the correlation between the qualitative and quantitative of cellulase activity. Data showed high positive correlation coefficient in enzyme activity in SSF and qualitative screening with (CMC & PASC) and the higher correlation showed with PACS and SSF rice straw was about (0.595^{**}) in β -Glucosidase. Our obtained results are in coherence with (Florencio et al., 2012) that studied the screening procedure using plates were compared with cellulase production under SSF. A correlation coefficient was correlation between the Congo red test and SSF, demonstrating that the two methodologies were in good agreement. Results showed that there are a negative correlation between SmF CMC substrate and PASC Qualitative screening in levels of β-Glucosidase and CMCase. Moreover, there were negative correlation in SmF (Avicel) substrate and CMC Qualitative screening in levels of FPase, CMCase and β-Glucosidase.



**Correlation is significant at the 0.01 level (2-tailed). *Correlation is significant at the 0.05 level (2-tailed).

Figure (3): Correlation coefficient between qualitative screening with (CMC & PASC) substrate and quantitative in SmF (Avicel &CMC) substrate and SSF (RS&WS) substrate.

Firstly, Using SmF production for high yield and high quality, but it can't be used in second generation biofuel. Moreover, SmF was very expensive (Figure 4). Secondly, SSF are producing low yield of enzymes that can be used in second generation biofuel. Therefore, we need to improve the isolates to obtain high yield of enzymes and releasing high amount of glucose in SSF. Different substrate showed higher enzymatic activity when compared on CMC substrate than the microcrystalline cellulose (Pirzadah *et al.*, 2014).

Detection of Cellulase genes of *Trichoderma* species

The existence of cellulase genes was shown in Figure (5). The cellulase genes, *Chb* gene encoding cellobiohydrolase 2 (CBH2) recorded in all *Trichoderma* isolates. Moreover, the *Bgl* gene which was encoding endo- β -glucanase (BGL1) in all *Trichoderma* isolates except two isolates MNF-MAS-*Tricho* 5 and 6.

We carried out PCR for detection of important cellulase genes (Cbh and *Bgl*) in the genomes of

the twenty-six *Trichoderma* isolates as shown in Figure (5). The primers utilized for amplification of *Cbh* gene yielded one band (230:270 bp) in all tested isolates (Figure 5a). Moreover, for Bgl, the PCR produced one band (420 bp) that appeared in all isolates except two isolate Named MNF-MAS-*Tricho6* and 7 (Figure 5b) in our result we found more DNA band (750bp) appeared in 9 isolate MNF-MAS-*Tricho* 2, MNF-MAS-*Tricho* 4, MNF-MAS-*Tricho* 7, MNF-MAS-*Tricho* 11, MNF-MAS-*Tricho* 13, MNF-MAS-*Tricho* 19, MNF-MAS-*Tricho* 20, MNF-MAS-*Tricho* 22, MNF-MAS-*Tricho* 23.

Genes encoding the cello-bio-hydrolase enzyme (CBHI), designated as *cbhI*, The CBH enzymes are known as the key component of cellulase system that display an exo-type of digesting biopolymers, and the major product of their action on cellulose was cellobiose (Teeri, 1997) Based on amino acid sequence similarity, the CBH enzymes can be grouped into glycoside hydrolase families: GH6, GH7, and GH48(β glucosidase I (BGLI) gene plays a major role in the conversion of randomly cleaved cello oligosaccharides into glucose.

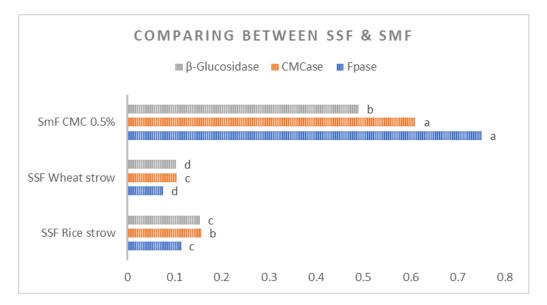
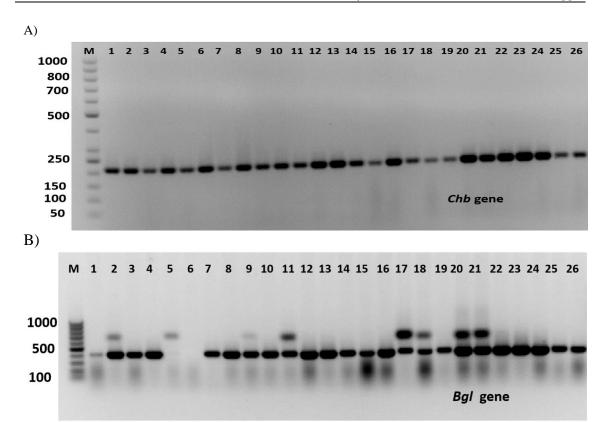


Figure (4): Comparison between different methods of enzyme production SSF (IU g-1) and SmF (IU ml-1)



Correlation between Qualitative and Quantitative Cellulase Enzymes activities in some *Trichoderma spp.*

Figure (5): Agarose gel analysis of specific-PCR products from amplification a:*Cbh* gene with about 230 bp of *Trichoderma* isolates. Lane 1: 50 bp DNA Ladder, B: *Bgl* gene with about 420 bp of *Trichoderma* isolates(Lane 1: 100 bp DNA Ladder).

Conclusion

In our study we were able to detect the correlation between Qualitative and quantitative characters of cellulase enzymes. Theise was a significant relationship between tested Trichoderma isolates. Those isolates were collected from various locations. In fact, we recommended usage this approach screening various Trichoderma isolates using PASC and CMC plate. This will facilitate identification of most promising high cellulases isolates.

REFERENCES

Alegre, A. C. P.; Polizeli, M. de L. T. de M.; Terenzi, H. F.; Jorge, J. A. and Guimarães, L. H. S. (2009). Production of thermostable invertases by Aspergillus caespitosus under submerged or solid state fermentation using agro-industrial residues as carbon source. *Brazilian J. Microbiol*, 40(3)612-622.

- Bose, R. G. (1963). A modified cellulosic medium for the isolation of cellulolytic fungi from infected materials and soils. *Nature*, *198*(4879).
- Castrillo, M. L.; Bich, G. Á.; Amerio, N. S.; Rodríguez, M. D., Zapata, P. D., & Villalba, L. L. (2021). Assessment of cellulase complex secretory capacity of *Trichoderma* strains and morphological and molecular identification of the isolate with the highest enzymatic secretion capacity. *Journal of Microbiology, Biotechnology and Food Sciences, 10*(5) 598-607
- Castrillo, M. L.; Bich, G. Á.; Modenutti, C.; Turjanski, A.; Darío Zapata, P. and Villalba, L. L. (2017). First Whole-Genome Shotgun Sequence of a Promising Cellulase Secretor, Trichoderma koningiopsis Strain POS7.
- Dewiyanti, I.; Darmawi, D.; Muchlisin, Z. A.; Helmi, T. Z.; Arisa, I. I.; Rahmiati, R.; Destri, E. and Fanisha, S. (2022). Characteristic and activity of cellulolytic bacteria isolated from mangrove soil in Northern Coast of Aceh

Province, Indonesia. *Biodiversitas*, 23(12): 6587-6599.

- Fahmi, A.I.; Eissa, R.A.; El-Halfawi, K.A.; Hamza, H.A. and Helwa, M. (2016). Identification of *Trichoderma* spp. by DNA Barcode and Screening for Cellulolytic Activity. *Journal of Microbial & Biochemical Technology*, 8(3): 202–209.
- Florencio, C.; Couri, S. and Farinas, C. S. (2012). Correlation between agar plate screening and solid-state fermentation for the prediction of cellulase production by *Trichoderma* strains. *Enzyme Research*, 2012.
- Ghose, T. K. (1987). Measurement of cellulase activities. Pure and Applied Chemistry, 59(2).
- González, R., Ramón, D., & Pérez-González, J. A. (1992). Cloning, sequence analysis and yeast expression of the egl1 gene from Trichoderma longibrachiatum. *Applied Microbiology and Biotechnology*, 38(3) 370-375.
- Gupta, N. (2017). First Report of Leek Yellow First Report Of Web Blight On. January, 20–21.
- Gupta, V. K.; O'Donovan, A.; Tuohy, M. G. and Sharma, G. D. (2014). Trichoderma in Bioenergy Research: An Overview. In Biotechnology and Biology of Trichoderma.
- Heng, J. L. S. and Hamzah, H. (2022). Effects of different parameters on cellulase production by Trichoderma harzianum TF2 using solid-state fermentation (SSF). *Indonesian Journal of Biotechnology*, 27(2): 80–86.
- Henrissat, B. (1991). A classification of glycosyl hydrolases based on amino acid sequence similarities. *Biochemical Journal*, 280(2): 781-8.
- Kazeem, M. O.; Ajijolakewu, K. A. and Rahman, N. A. A. (2021). Cellulase Production by Coculture of Bacillus licheniformis and B paralicheniformis over Monocultures on Microcrystalline Cellulose and Chicken Manure-supplemented Rice Bran Media. In BioResources (Vol. 16, Issue 4, pp. 6850-6869). North Carolina State University. https://doi.org/10.15376/biores.16.4.6850-6869
- Kshirsagar, S.; Waghmare, P.; Saratale, G.; Saratale,
 R.; Kurade, M.; Jeon, B. H. and Govindwar, S.
 (2020). Composition of Synthesized
 Cellulolytic Enzymes Varied with the Usage of
 Agricultural Substrates and Microorganisms.

Applied Biochemistry and Biotechnology, 191(4).

- Liu, Y. S.; Baker, J. O.; Zeng, Y.; Himmel, M. E.; Haas, T. and Ding, S. Y. (2011). Cellobiohydrolase hydrolyzes crystalline cellulose on hydrophobic faces. *Journal of Biological Chemistry*, 286(13).
- Elsobky, M. A.; Fahmi, A.; Ragaa, Eissa and Elzanaty, A. M. (2019). Genetic Characterization of Trichoderma spp. Isolated from Different Locations of Menoufia, Egypt and Assessment of their Antagonistic Ability. *J Microb Biochem Technol*, 11(1)9-23.
- Marecik, R.; Błaszczyk, L.; Biegańska-Marecik, R. and Piotrowska-Cyplik, A. (2018). Screening and identification of trichoderma strains isolated from natural habitats with potential to cellulose and xylan degrading enzymes production. *Polish Journal of Microbiology*, 67(2 181-190).
- Miller, G. L. (1959). Use of Dinitrosalicylic Acid Reagent for Determination of Reducing Sugar. *Analytical Chemistry*, 31(3) 426–428.
- Mueansichai, T.; Rangseesuriyachai, T.; Thongchul, and Assabumrungrat, N. S. (2022).Lignocellulosic Bioethanol Production of Napier Grass Using Trichoderma reesei and Saccharomyces cerevisiae Co-Culture Fermentation. International Journal of Renewable Energy Development, 11(2): 423-433.
- Naher, L.; Fatin, S. N.; Sheikh, M. A. H.; Azeez, L. A.; Siddiquee, S.; Zain, N. M. and Karim, S. M. R. (2021). Cellulase enzyme production from filamentous fungi trichoderma reesei and aspergillus awamori in submerged fermentation with rice straw. *Journal of Fungi*, 7(10): 1–11.
- Nathan, V. K.; Rani, M. E.; Rathinasamy, G.; Dhiraviam, K. N. and Jayavel, S. (2014). Process optimization and production kinetics for cellulase production by Trichoderma viride VKF3. SpringerPlus, 3(1)2-12.
- Oni, O. D.; Oke, M. A. and Sani, A. (2020). Mixing of Prosopis africana pods and corn cob exerts contrasting effects on the production and quality of Bacillus thuringiensis crude endoglucanase. *Preparative Biochemistry and Biotechnology*, 50(7) 1-10.
- Patel, A. K.; Pandey, A. and Singhania, R. R. (2019). Production of celluloytic enzymes for lignocellulosic biomass hydrolysis. In *Biomass*,

Correlation between Qualitative and Quantitative Cellulase Enzymes activities in some Trichoderma spp.

Biofuels, Biochemicals: Biofuels: Alternative Feedstocks and Conversion Processes for the Production of Liquid and Gaseous Biofuels.

- Pirzadah, T.; Garg, S.; Singh, J.; Vyas, A.; Kumar, M.; Gaur, N.; Bala, M.; Rehman, R.; Varma, A.; Kumar, V. and Kumar, M. (2014). Characterization of Actinomycetes and Trichoderma spp. for cellulase production utilizing crude substrates by response surface methodology. *SpringerPlus*, 3(1)2-12.
- Pointing, S. B. (1999). Qualitative methods for the determination of lignocellulolytic enzyme production by tropical fungi. *Fungal Diversity*, 2: 17–33.
- Rautela, G. S. and Cowling, E. B. (1966). Simple Cultural Test for Relative Cellulolytic Activity of Fungi. *Applied Microbiology*, 14(6): 892-8.
- Sazci, A.; Erenler, K. and Radford, A. (1986). Detection of cellulolytic fungi by using Congo red as an indicator: a comparative study with the dinitrosalicyclic acid reagent method. *Journal of Applied Bacteriology*, 61(6): 559 - 562.
- Shad, M.; Hussain, N.; Usman, M.; Akhtar, M. W. and Sajjad, M. (2023). Exploration of computational approaches to predict the structural features and recent trends in αamylase production for industrial applications. In *Biotechnology and Bioengineering*, 120 (8) 2092.
- Sharrock, K. R. (1988). Cellulase assay methods: a review. In *Journal of Biochemical and Biophysical Methods*, 17(2).
- Suirta, I. W.; Suarsana, I. N.; Swantara, I. M. D. and Proborini, M. W. (2021). Morphological and molecular characterization of the pathogenic fungi isolated from purple eggplant originating

from bali, indonesia. *Sabrao Journal of Breeding and Genetics*, 53(3): 459-467.

- Teather, R. M. and Wood, P. J. (1982). Use of Congo red-polysaccharide interactions in enumeration and characterization of cellulolytic bacteria from the bovine rumen. *Applied and Environmental Microbiology*, 43(4): 777-80.
- Teeri, T. T. (1997). Crystalline cellulose degradation: New insight into the function of cellobiohydrolases. *Trends in Biotechnology*, *15*(5).
- Wang, J. L. and Gao, P. J. (1999). PCR-mediated analysis of transcription of CBH I and II genes from Trichoderma pseudokoningii and Penicillium janthinellum. In *Biotechnology Letters* (Vol. 21).
- Yadav, R.; Kumar, V.; Baweja, M. and Shukla, P. (2018). Gene editing and genetic engineering approaches for advanced probiotics : A review. *Critical Reviews in Food Science and Nutrition*, 58(10): 1735–1746.
- Zahra, T.; Irfan, M.; Nadeem, M.; Ghazanfar, M.; Ahmad, Q.; Ali, S.; Siddique, F.; Yasmeen, Z. and Franco, M. (2020). Cellulase Production by Trichoderma viride in Submerged Fermentation using Response Surface Methodology. *Punjab* University Journal of Zoology, 35(2): 27-33.
- Zeng, W., & Chen, H. Z. (2009). Air pressure pulsation solid state fermentation of feruloyl esterase by Aspergillus niger. *Bioresource Technology*, 100(3).
- Zhang, J., Zhang, G., Wang, W., Wang, W., & Wei, D. (2018). Enhanced cellulase production in Trichoderma reesei RUT C30 via constitution of minimal transcriptional activators. *Microbial Cell Factories*, 17(1): 2-14.

دراسات مقارنة لنشاط إنزيمات السليوليز في الاختبارات الوصفية والكمية فى فطر الترايكوديرما

محمد علاء الدين محمد السبكي، خالد صلاح الدين محمد، عبد المجيد إبراهيم فهمي، عبد الفتاح مندي الزناتي، رجاء عبد العزيز عيسى قسم الوراثة ، كلية الزراعة جامعة المنوفية

الملخص العربى

تم دراسة سنة و عشرون عزلة من فطر الترايكوديرما من خلال الاختبارات الوصفية لأنزيم السيليوليز من خلال اطباق CMC مع صبغة الكونجو الأحمر و كذلك اطباق PASC. بالإضافة الي الاختبارات الكمية من خلال التخمرات الصلبة والسائلة. كما تم عمل التخمرات السائلة علي بيئة ماندل مع PASC. من CMC تم قياس الانزيمات -β (FPase, CMCase , β- ما النزيمات -β (Pase, CMCase , β- ما النزيمات -β (FPase, CMCase , β- و تم قياس كفاءة هذه الانزيمات علي البيئات الصلبة علي قش الأرز و السائلة. كما تم عمل التخمرات السائلة علي بيئة ماندل مع PASC ، و CMC تم قياس الانزيمات -β (FPase, CMCase , β- و تم قياس كفاءة هذه الانزيمات علي البيئات الصلبة علي قش الأرز و القمح. و القمح. و تم قياس كفاءة هذه الانزيمات علي البيئات الصلبة علي قش الأرز و القمح. و القمح. و المعنوية الي درجة كبير عند و ٩٠ و و ١ ك و تم قياس كفاءة هذه الانزيمات علي البيئات الصلبة علي قش الأرز و القمح. و القمح. و القمح. و المعنوية الي درجة كبير عند و ٩٠ و ٩٠ و الكمية في التخمرات السائلة و الصلبة و الصلبة و الصلبة و الما وسفي و الكمي موجب المعنوية الي درجة كبير عند و٩٠ و٩٠ و الكمية في التخمرات السائلة و الصلبة و الصلبة و الما ومني الوصفي و الكمي موجب المعنوية الي درجة كبير عند و٩٠ و٩٠ والكمية في التخمرات السائلة و الصلبة وي قش الأرز. و بالمقارنة بين طرق إنتاج الانزيمات علي البيئة الصلبة و السائلة اتضح ان انتاج الانزيم من التخمرات السائلة تعطى كمية اكثر كما إنها بين طرق إنتاج الانزيمات الما الما وكثلك الما والنه الحمرات السائلة تعلى كما الما وي الما وكثل كا السلالات المدروسة يوجد بها هذه الجينات .